The investigation of adrenal involvement in carbapenem resistant Acinetobacter baumannii sepsis: experimental mouse model

Abstract

Background/aim: In the last years, incidence of carbapenem resistant Acinetobacter baumannii sepsis is increasing with high mortality. However, it is not clear whether this is due to inadequate antimicrobial choice or a more severe clinical course. We aimed to evaluate the inflammation and adrenal involvement in the carbapenem resistant A. baumannii by using experimental mouse model sepsis.

Materials and methods: Balb/c female mice were randomly put into control and three sepsis groups (A. baumannii susceptible to carbapenem-CSAB-, A. baumannii resistant to carbapenem-CRAB-, Escherichia coli). A total of sixty mice were included in this study with each group having 15 mice. Mice were sacrificed, 72 hours after bacterial inoculation and blood was taken from each mouse for the assessment of cytokines and corticosterone. Both adrenal glands were dissected; one was used for culture and the other was used for histopathological examination. Bacterial loads of organs were calculated as CFU/g. The histopathological changes, bacterial levels in adrenal and cytokine and corticosterone levels were assessed and compared among the groups.

Results: The bacterial level was higher in E. coli (108, 45 ±30, 55 log10 CFU/g) (mean±SD) than other sepsis groups. The lowest level of corticosterone was observed in the E. coli group (p<0.001). TNF alpha level was highest in the CRAB and E. coli group and this difference was statistically significant than control group (p<0.05). The IL-6 level in CRAB was significantly higher than the control group (10, 20 pg/ml). The adrenal gland congestion was significantly severe in all the sepsis groups compared to the control.
In the group comparison, congestion was significantly more severe in the *E. coli* group than in CSAB and CRAB groups.

**Conclusion:** Adrenal involvement and inflammatory reactions are seen in *E. coli* sepsis and in CRAB sepsis. These findings will be focused on in future clinical trials.

**Key words:** *Acinetobacter baumannii*, adrenal insufficiency, corticosteroids, mice model, sepsis

1. **Introduction**

Sepsis is defined as a life-threatening organ dysfunction caused by dysregulated host response to infection [1]. It is an inflammatory disease that can cause impairment of tissue perfusion, multiple organ failure, shock and even death. Sepsis is defined by the World Health Organization (WHO) as a global health problem and the reported mortality rate of sepsis in hospitalized patients ranges from 30 to 45%. Despite recent advances in treatment and technology, the sepsis mortality rate is still relatively high. Although sepsis pathogenesis is not clearly understood, it is known that antigens and toxins of microorganisms induce inflammation. Many systems and cells are activated by bacterial products such as antigens, toxins, enzymes etc. and lead to pathophysiological events such as the release of cytokines, neutrophil migration through chemotaxis, impairment of endothelium permeability, hyperactivation of coagulation pathway. These events can result in perfusion impairment, hypoxia, edema and necrosis. These had to organ dysfunction and/or organ failure that consists of the central nervous system, lung, kidney, liver and/or hemopoietic systems [2, 3].

In cases of increased stress in the body such as trauma, pain or inflammation, the hypothalamo-hypophyseal adrenal axis (HPA) is activated. The level of glucocorticoid increases with the release of adrenocorticotropic hormone (ACTH) stimulus [4, 5]. It is
known that the adrenal glands are affected by certain infections; such as meningococcemia, brucellosis, tularemia etc. HPA is affected by the direct invasion of microorganisms into the organs or due to toxin effect during sepsis. Adrenal failure seems to be one of the most important prognostic factors in the outcome of sepsis. Adrenal insufficiency leads to inadequate stress response and may increase mortality. Currently, adrenal insufficiency is still a debatable issue [6].

According to the recent treatment guidelines, hydrocortisone treatment is recommended (200 mg / day) for patient with septic shock, who do not respond adequately to fluid replacement and vasopressor treatment [1]. However, the effect of steroid treatment on reducing mortality in sepsis is still controversial [3]. In previous clinical study with sepsis, basal cortisol level was higher in the patients with shock than without shock. Basal cortisol level was also lower in the patients with gram-negative bacterial sepsis than in the patients with gram-positive bacterial sepsis [7]. In recent years, the incidence of nosocomial sepsis has been increased due to multi drug resistant (MDR) bacteria [8]. One of the most common gram-negative bacteria is MDR Acinetobacter baumannii in the intensive care unit (ICU). According to Xx University’s Hospital Infection Control Surveillance report (2016), A. baumannii is the most common cause of ICU-acquired infections [9]. It is also known that MDR A. baumannii has a higher mortality than antibiotic susceptible A. baumannii. However, it is not clear whether or not this is due to an inadequate choice of antimicrobials or if there is a more severe clinical reason [10, 11]. The aim of this study is to evaluate whether or not the inflammation and adrenal involvement in the carbapenem resistant A. baumannii differs from carbapenem susceptible gram-negative bacteria in experimental mouse model sepsis.

2. Materials and methods
This experiment was carried out at the Experimental and Clinical Research Center at xx. The study was approved the local ethics committee for animal experimentation.

2.1. Animal

Balb/c female mice aged 20-24 weeks weighing 40-50 g were used in the study. All mice were caged in groups of three or four and given ad libitum access to food and water. Animals were acclimatized in a 12-hour light/dark cycle.

2.2. Bacteria

The study was performed with strains of carbapenem susceptible A. baumannii (CSAB) ATCC 17978, carbapenem resistant A. baumannii (OXA-51, OXA-58, PER-1 positive; CRAB) and E. coli ATCC 25922. CRAB was a clinical isolate obtained from the blood culture of a bacteremic patient at the hospital.

2.3. In vivo study

The mice were randomly put into two main groups; control group and sepsis group. The sepsis groups were also divided into three subgroups; CSAB, CRAB and E. coli study group. A total of sixty mice were included in this study. The control group and sepsis subgroups each consisted of 15 mice. Sepsis was developed by an intraperitoneal injection (ip) of 0.1 ml bacterial suspensions, which had been determined for each group in the preliminary study (Table 1). All mice were sacrificed at 72 h with an overdose of anesthetic (100 mg/kg ketamine hydrochloride, Pfizer, Turkey) and 10 mg/kg xylazin (Xylazin bio %2®, 20 mg/ml flakon, Bioveta). Intracardiac blood samples were taken for cytokine analysis and corticosterone. Tissue samples were taken from the lung, liver, heart and right adrenal gland and cultured on tryptic soy agar (Merck, Germany) to check the development of sepsis. Sepsis was defined with observation of the bacterial growth at least two organs excluding peritoneum. Quantitative culture was performed and results
were given as CFU per gram for adrenal gland [10,11]. Also, 10-fold dilutions of right adrenal glands homogenates were processed to obtain quantitative counts as CFU/g would be used to compare the control and study groups. The left adrenal glands were used for histopathological examination.

2.4. Cytokine and corticosterone analysis

Serum samples were stored at -20 °C for corticosterone, TNF-alpha and IL-6 according to the kit procedure instructions using enzyme-linked immunosorbent assay (ELISA) method (FineTest, Mouse CORT (Corticosterone) ELISA Kit EU3108, Mouse TNF-alpha (Tumor Necrosis Factor Alpha) ELISA Kit EM0183, Mouse IL-6 (Interleukin 6) ELISA Kit EM0121) The cytokine levels were measured by ELISA plate reader (GloMax® Microplate Reader, PROMEGA).

2.5. Histopathological examination

The left adrenal glands from all the mice were fixed in 10% neutral buffered formalin solution. After 24 hours of fixation, all tissues were blocked in paraffin and 5 micrometer sections. They were prepared and stained with hematoxylin eosin following by being examined under light microscope. In the microscopic examination, three layers of the cortex and the adrenal glands medulla were evaluated separately. All layers of the adrenal gland were evaluated for congestion, neutrophil infiltration and other possible changes. The changes were recorded semi-quantitatively as 0: none, 1: mild, 2: moderate, 3: severe to allow statistical comparisons (Table 2). Pictures are given as an example of the evaluation in Figure.

2.6. Statistical analysis

Before the study, a power analysis was studied. The results of the power analysis showed that 15 mice per group were required to achieve 80% power. Bacterial counts from
quantitative cultures of adrenal gland tissue, serum corticosterone, IL-6 and TNF-alpha levels and tissue histological grades according to the groups were compared and analyzed using SPSS 22.0 (SPSS Inc., Chicago, IL, USA). Results of the microbiological study were recorded as log10. Shapiro Wilk test for normality distribution analysis was performed before analysis. A One-way ANOVA was used to test for differences among the groups. A post-hoc Tukey test was performed to determine which group had a different result. Data were presented as mean ± standard deviation. A p-value of <0.05 was considered significant statistically.

3. Results

This study involved 60 mice, but 2 mice were excluded from the CRAB group due to death within 24 hours (Table 1).

3.1. Microbiological examination

Sepsis developed in all study groups. Bacterial growth in organ cultures are shown in Table 3. The bacterial levels of adrenal glands were higher in *E. coli* than CRAB and CSAB groups. The differences were proven to be statistically significant (p<0.001).

3.2. Histopathological examination

In the microscopic examination, normal adrenal tissue was observed in the control group (Figure A). Neutrophil leukocyte infiltration was not observed in the sepsis groups. However, congestion of the adrenal gland was detected in the groups (Figure B). The adrenal gland congestion was significantly more severe in the sepsis groups compare to the control group (Table 3). In the comparison of the sepsis groups, congestion was significantly more severe in the *E. coli* than in CSAB and CRAB groups (p <0.05). The histopathological examination, vacuolated cells between the deep parts of the cortex and the medulla were observed in some adrenal glands (Figure C and D). These cells were
also graded and recorded. Vacuolated cells were observed in 6 mice in the CSAB group, 12 mice in the CRAB group, and 14 mice in the *E. coli* group however vacuolated cells were not present in the control group. There were more vacuolated cells in the CRAB and *E. coli* group than in the control and CSAB groups. Statistical analysis showed that the difference between the control group of two sepsis groups (CRAB and *E. coli* groups) was significant (p<0.001).

### 3.3. Hormone and Cytokine Assays

The corticosterone levels were lower in the *E. coli* group than any other sepsis groups and the control group. Also, corticosterone level was lower in the CRAB than CSAB (Table 3). The difference was found to be statistically significant (p<0.05).

The highest TNF-alpha mean level was in the CRAB group (Table 3). The mean of TNF-alpha level was significantly higher in CRAB and *E. coli* than the control group (p <0.05).

However, this difference was not statistically significant.

CRAB group had the highest IL-6 level (Table 3). In statistical analysis only, IL-6 mean level in the CRAB group was statistically significantly higher than the control group (p = 0.009).

### 4. Discussion

Throughout the world, severe sepsis and septic shock still remain major health problem. Despite improvements in medical care, severe sepsis is still associated with high mortality [12]. Rapid activation of the adrenal glands glucocorticoid and catecholamine production is a fundamental component of the stress response and is essential for survival of the host. Multi-organs including adrenal glands are also affected in sepsis, leads to organ dysfunctions. In these patients, the plasma levels of ACTH and cortisol are often affected. The HPA axis and glucocorticoid actions are shown to be impaired in many critically ill
patients. In previous studies the adrenal insufficiency rate has been reported to vary between 10 to 20% and may be as high as 60% in cases with septic shock [13]. It is not known which bacterial agent has more influence on the adrenal glands. However, certain agents such as Neisseria meningitis, Mycobacterium tuberculosis are known to affect adrenal glands [14].

Many different microorganisms are responsible for adrenal involvement in sepsis. In bacterial infections, many pathological conditions such as massive bleeding, abscess and granulomas are observed. Hemorrhage is the most common lesion in adrenal. There is also a correlation between the presence of bacterial involvement in the adrenal gland and adrenal hemorrhage [15]. The current study, histopathological examination of the adrenal glands revealed no leukocytes, hemorrhages and necrosis in any of the layers. However, congestion was observed in different layers among the groups. Profuse congestion was observed in the *E. coli* and CRAB groups, whereas less severe congestion was observed in CSAB group. The difference in the effect on the adrenal glands and the severity of sepsis between mice infected with CSAB or CRAB strain cannot be contributed to carbapenem resistance and due to the difference in the virulence of the strains.

It has been reported in the literature that vacuolated cells in corticomedullary complements of the adrenal glands are present in mice and disappear with puberty reputation. These cells are not seen in human adrenals*. The presence of these cells in

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mouse studies is accepted as normal. The current study showed that the vacuolated cells increased in the severe sepsis groups. This may suggest the effect of stress on adrenal glands. More studies are needed to study the effect of stress on adrenal glands as well as. Hormonal levels should be studied from these vacuolated cells.

Marik et al. found that 25% of patients in 59 sepsis cases in their study had adrenal insufficiency and 17% had hypothalamo-hypophyseal axillary failure [16]. In another study that consisted of in 189 severe sepsis patients, adrenal insufficiency was observed in 10% of patients. Adrenal insufficiency is correlated with increasing mortality [17]. The corticosterone level was significantly lower in *E. coli* group. In addition, corticosterone levels in the CRAB group were lower than in the CSAB group. This suggests that adrenal insufficiency develops in the sepsis of *E. coli* and CRAB.

Adrenal functions are related to inflammation. Endotoxemia increases circulating IL-1, IL-6 and TNF-a, and these cytokines acutely and transiently activate HPA; thus, increasing the release of cortisol [18]. On the other hand, products such as TNF-a and corticostatin can inhibit the production of adrenal function and cortisol [19, 20]. In particular, inflammatory mediators have been shown to have suppressive effects on adrenal function. Prolonged exposure to cytokines may result in an altered response of the hypothalamic-pituitary axis. Similarly, the chronic increase in IL-6 may lead to a reduction in ACTH production, it has been shown that TNF-a may cause a reduction in corticotrophin releasing hormone (CRH) stimulation and ACTH production in the adrenal function [21, 22]. Low ACTH levels have been observed in patients that were diagnosed with severe sepsis or systemic inflammatory response syndrome [18]. Patients with severe sepsis were evaluated and found to have lower mean cortisol levels in the group with higher mortality in a study [22]. Obviously the HPA axis relation with sepsis is not clearly
defined. In the inflammatory response, there is some evidence that cortisol production increases in patients, but nearly half of the patients with septic shock have an inadequate response to metyrapone synthesis which suggests a reduction in cortisol synthesis [23]. In our study, IL-6 and TNF-α levels were high in the *E. coli* and CRAB group where the cortisol level was measured at the lowest level in the *E. coli* group. The bacterial level in the adrenal gland was significantly higher in the *E. coli* group whereas the level of corticosterone was lower in *E. coli* and CRAB (Table 3).

Several studies on septic shock have reported that IL-6 and TNF-α levels are elevated and have a correlation between cytokines levels cytokines and mortality [18]. In another study, high TNF-α levels were reported as a high predictive value for mortality for gram-negative sepsis [24]. In the presence of inflammation, IL-6 has an enhancing effect on serum corticotropin and cortisol levels [19]. Andaluz-Ojeda et al. in a sepsis study with 17 immunomodulators, found that IL-6 levels were correlated with mortality rates between 3 and 28 days. Wu et al. found that there were higher levels of IL-6 in septic shock patients when compared to non-shock sepsis patients and group with higher mortality [25, 26]. In our study, TNF alpha levels were significantly higher in the *E. coli* and CRAB groups compared to the control group, and the highest TNF alpha level was detected in the CRAB group. IL-6 levels were higher in three of the sepsis groups than in the control group. Statistically, only the CRAB group was statistically significantly different from the control group.

In this experimental mouse model sepsis, corticosterone levels were lower in the CRAB group than CSAB group. Also, cytokine levels (TNF alpha and IL-6) were highest in the CRAB sepsis than CSAB sepsis. In conclusion, a similarity adrenal involvement and
inflammatory reactions are seen in E. coli sepsis and in CRAB sepsis which have high
virulence.

In this study, 15 mice in each group were determined by power analysis. However, it is
an important limitation that two mice in the CRAB group were not included in the study
because they died of sepsis. Further studies with larger working groups are needed. The
authors believe that these findings will shed light on future clinical trials.

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10.1007/s00011-009-0003-0
Table 1. Study groups for the experimentation

<table>
<thead>
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<th>Group</th>
<th>n</th>
<th>Inoculum dose</th>
<th>72 h</th>
</tr>
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<tbody>
<tr>
<td>Control Group</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Group 1</td>
<td>15</td>
<td>Serum Physiologic</td>
<td>Sacrification</td>
</tr>
<tr>
<td>Sepsis Groups</td>
<td></td>
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<tr>
<td>Group 2</td>
<td>15</td>
<td>4x10^8 cfu/ml (0.1 ml)</td>
<td>Sacrification</td>
</tr>
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<td>CSAB</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Group 3</td>
<td>13*</td>
<td>3x10^8 cfu/ml (0.1 ml)</td>
<td>Sacrification</td>
</tr>
<tr>
<td>CRAB</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Group 4</td>
<td>15</td>
<td>1x10^8 cfu/ml (0.1 ml)</td>
<td>Sacrification</td>
</tr>
<tr>
<td>E. coli</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

* Two mice were excluded from the study died within 24 hours as they.

CSAB: Carbapenem susceptible *A. baumannii*.

CRAB: Carbapenem resistant *A. baumannii*. 
Table 2. Scores of histopathological examination

<table>
<thead>
<tr>
<th></th>
<th>None</th>
<th>Mild</th>
<th>Moderate</th>
<th>Severe</th>
</tr>
</thead>
<tbody>
<tr>
<td>Congestion</td>
<td>0</td>
<td>1</td>
<td>2</td>
<td>3</td>
</tr>
<tr>
<td>Neutrophil infiltration</td>
<td>0</td>
<td>1</td>
<td>2</td>
<td>3</td>
</tr>
<tr>
<td>Vacuolated cells</td>
<td>0</td>
<td>1</td>
<td>2</td>
<td>3</td>
</tr>
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</table>
### Table 3. Bacterial load and congestion in adrenal tissue, corticosterone and cytokines levels in the groups

<table>
<thead>
<tr>
<th>PARAMETERS</th>
<th>GROUPS</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Control (n=15)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>(n=15)</td>
<td></td>
</tr>
<tr>
<td>Bacterial count, log10 CFU/g</td>
<td>NG</td>
<td></td>
</tr>
<tr>
<td>Mean ±SD</td>
<td>71,46±26,12</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td></td>
<td>55,67±31,86</td>
<td></td>
</tr>
<tr>
<td></td>
<td>108,45 ±30,55</td>
<td></td>
</tr>
<tr>
<td>Corticosterone</td>
<td>76,66±3,31</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Mean ±SD (µg/ml)</td>
<td>80,63±4,22</td>
<td></td>
</tr>
<tr>
<td></td>
<td>71,96±4,87</td>
<td></td>
</tr>
<tr>
<td></td>
<td>62,71±9,96</td>
<td></td>
</tr>
<tr>
<td>TNF alpha</td>
<td>195,84±46,48</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Mean ±SD (pg/ml)</td>
<td>217,07±33,04</td>
<td></td>
</tr>
<tr>
<td></td>
<td>267,07±31,55</td>
<td></td>
</tr>
<tr>
<td></td>
<td>250,62±73,20</td>
<td></td>
</tr>
<tr>
<td>IL-6</td>
<td>7,24±2,62</td>
<td>0.010</td>
</tr>
<tr>
<td>Mean ±SD (pg/ml)</td>
<td>8,77±2,41</td>
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</tr>
<tr>
<td></td>
<td>10,20±2,36</td>
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</tr>
<tr>
<td></td>
<td>9,52±1,98</td>
<td></td>
</tr>
<tr>
<td>Congestion</td>
<td>0,46±0,63</td>
<td>0.009</td>
</tr>
<tr>
<td>Mean ±SD</td>
<td>4,30±1,70</td>
<td></td>
</tr>
<tr>
<td></td>
<td>3,73±2,08</td>
<td></td>
</tr>
<tr>
<td></td>
<td>6,00±1,64</td>
<td></td>
</tr>
<tr>
<td>Bacterial growth in liver</td>
<td>NG</td>
<td>10/15</td>
</tr>
<tr>
<td>---------------------------</td>
<td>----</td>
<td>-------</td>
</tr>
<tr>
<td>Bacterial growth in lung</td>
<td>NG</td>
<td>12/15</td>
</tr>
<tr>
<td>Bacterial growth in heart</td>
<td>NG</td>
<td>10/15</td>
</tr>
</tbody>
</table>

**Abbreviations and explanations**

1. SD: Standard deviation
2. NG: No Growth
3. CSAB: Carbapenem Susceptible *A. baumannii*
4. CRAB: Carbapenem Resistant *A. baumannii*
5. \(a\): \(p=0.001\) with CSAB, \(p<0.001\) with CRAB and control
6. \(b\): \(p=0.003\) with CSAB, \(c\): \(p=0.001\) with CRAB, \(p<0.001\) with CSAB and control
7. \(d\): \(p<0.001\) with control, \(e\): \(p=0.004\) with control
8. \(f\): \(p=0.009\) with control
9. \(g\): \(p<0.001\) with control, \(h\): \(p<0.001\) with control \(i\): \(p=0.036\) with CRAB, \(p=0.002\) with CSAB, \(p<0.001\) with control
10. \(j\): \(p<0.001\)
11. \(k\): \(p=0.030\), \(l\): \(p<0.001\)
12. \(m\): \(p<0.001\)
Figure (HE: Hematoxylin eosin)

A: Normal adrenal tissue (HEx40);
B: Septic adrenal tissue; Mild congestion (HEx40);
C: Septic adrenal tissue; severe congestion and fatty cell accumulation and in medulla, (HEx40);
D: Septic Adrenal Tissue severe congestion and fatty cell accumulation and in medulla (HEx100)