

1 **Investigation of the relationship between immune checkpoints and mismatch**
2 **repair deficiency in recurrent and non-recurrent glioblastoma**

3 **Abstract**

4 **Background/aim:** Microsatellite instability tests and programmed cell death-1 (PD-1) /
5 programmed cell death ligand-1 (PD-L1) in the immune checkpoint pathway are the tests
6 that determine who will benefit from immune checkpoint inhibitor therapy. We aimed to
7 show the expression of DNA mismatch repair proteins and PD-1 / PD-L1 molecules that
8 inhibit immune checkpoints, to explain the relationship between them, and to demonstrate
9 their predictive role in recurrent and non-recurrent glioblastoma.

10 **Materials and methods:** We analyzed 27 recurrent and 47 non-recurrent cases at our
11 archive. We performed immunohistochemical analysis to determine expressions of PD-
12 1, PD-L1 and mismatch repair proteins in glioblastoma. We evaluated the relationship
13 between these two group and compared the results with the clinicopathological features.

14 **Results:** The mean age of diagnosis was significantly lower in recurrent glioblastoma
15 patients. Median survival was longer in this group. We found that PD-L1 expression was
16 reduced in recurrent cases. Additionally, recurrent cases had a significantly higher rate of
17 microsatellite instability. Loss of PMS2 was high in both group but was substantially
18 higher in recurrent cases.

19 **Conclusion:** The presence of microsatellite instability and low PD-L1 levels, which
20 are among the causes of treatment resistance in glioblastoma, were found to be
21 compatible with the literature in our study, with higher rates in recurrent cases. In
22 recurrent cases with higher mutations and where immunotherapy resistance is expected
23 less, low PD-L1 levels thought that different combinations with other immune

1 checkpoint inhibitors can be tried as predictive and prognostic marker in GBM
2 patients.

3 **Key words:** Glioblastoma, mismatch repair, programmed cell death protein ligand 1 (PD-
4 L1), programmed cell death protein 1 (PD-1), microsatellite instability

5 **1. Introduction**

6 Glioblastoma (GBM) is the most common primary brain tumor with a poor prognosis
7 (47.1%) [1]. GBMs are fatal tumors with a median survival time of 12 months.
8 Approximately 3-5% of cases live more than 3 years [2]. Many studies are showing the
9 importance of genetic susceptibility, exogenous factors, age and clinical parameters at the
10 time of diagnosis, as the reason for aggressive behavior [3-6]. Other important factors are
11 surgical procedures and multimodal treatment options. Immunotherapy is a promising
12 treatment method that shows a synergistic effect with radiotherapy [7].

13 Immune checkpoints can be grouped into two main groups as immunostimulating and
14 blocking. The main molecules that inhibit immune checkpoints are the Cytotoxic T-
15 Lymphocyte-Associated protein-4 (CTLA-4), Programmed Death-1 (PD-1) receptor, and
16 its two ligands (PD-L1 and PD-L2). These molecules block control signals that lead to
17 the T cell response against the tumor. New treatments are aimed at inhibiting PD-L1 on
18 the tumor cell or PD-1 receptor on the T cell which will produce an anti-tumoral response.
19 These treatment agents are called immune checkpoint inhibitors (ICIs). CTLA-4, one of
20 the major inhibitory molecules in GBMs, is released from T cells, binding to its ligands
21 (CD80, CD86) reduces the activation and proliferation of effector T cells and increases
22 the activation of regulatory T cells (Treg) in the GBM microenvironment [8,9].

23 PD-1 is released from T cells and other immune cells. One of its ligands, PD-L1 is found
24 in Tregs in the GBM microenvironment, tumor-associated macrophages, and other cells

1 in the tumor microenvironment, including tumor cells [8,10]. The predictive markers in
2 PD-1/PD-L1 antibody therapy are mainly the number of cytotoxic T-lymphocytes inside
3 tumor tissues and the expression level of PD-L1 in cancer cells [11,12]. In extensive
4 studies, PD-L1 expression levels in GBMs were found between 61-88% [13-15]. High
5 PD-L1 mRNA expression level was found to be associated with shorter overall survival
6 in glioma patients [12,16,17].

7 Treating recurrent GBMs is more difficult and resistance develops more frequently to
8 treatment. These tumors have been treated several times (radiotherapy and
9 temozolomide) and therefore contain a greater number of mutations. Due to the presence
10 of potential new antigens, these tumors are assumed to be more suitable for the
11 recognition and attack of the immune system. In contrast to the hypothesis that the tumor
12 assumes that it will increase the release of PD-L1 with the immune-escape mechanism
13 developed to protect itself from stronger immune response, in some studies, this rate was
14 found to be significantly lower in recurrent tumors [13,14,18].

15 PD-1 / PD-L1 expression, tumor mutation load, and DNA mismatch repair (MMR)
16 defects are thought to be related to the treatment response. Some clinical studies revealed
17 that defective MMR is associated with clinical responses to immune checkpoint inhibitors
18 (ICI) [5,19]. Tumors with high microsatellite instability (MSI) and high immunogenicity
19 benefit from immunotherapy more and have a better clinical course. Therefore, it is
20 recommended to use MSI status as a marker for response to PD-1 / PD-L1 blockade in
21 cancer patients [20].

22 In the last 6 years, significant results have been obtained in immunotherapy in various
23 tumors (melanoma, renal cell cancer, lung cancer, head and neck cancers) with anti-PD-
24 1 / PD-L1 antibodies. While the response to PD-1 inhibitors is significantly high in

1 lymphoma subtypes (87% in Hodgkin's lymphoma), this rate ranges from 15 to 40% in
2 solid organ cancers [21,22]. There are several studies showing that PD-1 inhibition
3 increases the antitumor responses and survival rate on animal glioma models [23,15].
4 Especially combined immune checkpoint blockade resulted in 100% long survivors [23].
5 However, there is insufficient clinical evidence to support its effectiveness in GBM
6 patients. There are some "case reports" in the literature showing that anti-PD-1 therapy
7 (nivolumab) have significant therapeutic effects on GBM patients [19,24,25].

8 **2. Material and methods**

9 **2.1 Patients and Clinical Information**

10 In our study, 74 cases (recurrent and non-recurrent GBM) diagnosed in our department
11 between 2007-2019 were selected. Twenty-seven of these cases showed recurrence.
12 Hypercellular tumors including palisading necrosis and vascular endothelial proliferation
13 were accepted as 'original tumor'. Diffuse necrosis, vascular hyalinization, gliosis, and
14 the presence of rare atypical cells were accepted as 'radiotherapy effect'. High-grade
15 glioma with mitosis and minimal evidence of radiation effect is defined as 'recurrent
16 tumor' [26]. The first resection materials of the recurrent cases were defined as "Group
17 1", the 2nd resection materials were "Group 2" and the non-recurrent cases were defined
18 as "Group 3". When calculating rates in recurrent cases, cases were considered "positive"
19 if any of the cases in group 1 or 2 were positive. The results of recurrent and non-recurrent
20 GBM patients were compared. In addition, in recurrent cases, the expression rates in the
21 1st and 2nd resection materials were compared. For each case, the patient's age at the time
22 of diagnosis, sex, time for recurrence, and survival were recorded. Clinical information
23 was obtained from patient files on the computer.

24 **2.2 Immunohistochemical Study**

1 Immunohistochemically, the relationship between MLH1 (MutL Homolog 1), MSH2
2 (MutS Homolog 2), MSH6 (MutS Homolog 6), and PMS2 (PostMeiotic Segregation
3 increased 2) results and immune checkpoint inhibitors PD-1 and PD-L1 were examined
4 and compared with the clinicopathological features. Sections stained with
5 immunohistochemical antibodies were examined under a light microscope (Olympus
6 BX50). Normal colon tissue was used as a control of the immunohistochemical MLH1,
7 MSH2, MSH6, and PMS2 markers. Expressions of MMR proteins were evaluated as
8 follows : Nuclear staining in more than 80% neoplastic cells was accepted as score 4;
9 51-80% staining score 3; 10-50% staining score 2; less than 10% staining score 1 and
10 no nuclear staining score 0 [27]. Expressions of MMR proteins were shown in Figure 1
11 (Figure 1: a) Loss of nuclear staining with MLH1 ($\times 200$), b) Nuclear staining with
12 MLH1 ($\times 100$), c) Loss of nuclear staining with MSH2 ($\times 200$), d) Nuclear staining with
13 MSH2 ($\times 100$), e) Loss of nuclear staining with MSH6 ($\times 200$), f) Nuclear staining with
14 with MSH6 ($\times 200$), g) Loss of nuclear staining with with PMS2 ($\times 100$), and h) Nuclear
15 staining with PMS2 ($\times 100$)). Loss of nuclear expression of one or more MMR proteins
16 was accepted as deficient mismatch repair [28].

17 SP263 clone of the PD-L1 antibody was used. Diffuse fibrillar/membranous staining in
18 tumor tissue was evaluated. According to previous studies, no staining in non-necrotic
19 tumor tissue was accepted as score 0; <25% staining score 1; 25-50% staining score 2;
20 50-75% staining score 3 and > 75% staining score 4 [13]. Membranous staining in
21 epitheloid tumor cells was defined as (+) if > 5% staining in tumor cells as in previous
22 studies [13].

23 PD-1 expression was seen in lymphocytes in tumor tissue, and perivascular space. It
24 was scored as sparsely, moderately and intensively according to the staining rates in
25 large magnification (200 \times -400 \times). The staining patterns of PD-L1 and PD-1 were shown

1 in Figure 2 (Figure 2: a) Diffuse fibrillary PD-L1 staining in tumor matrix ($\times 200$), b)
2 Membranous PD-L1 staining on tumor cells ($\times 400$), c) Diffuse fibrillary and
3 membranous PD-L1 staining ($\times 200$), d) PD-1 staining on tumor infiltrating lymphocytes
4 ($\times 400$)).

5 **2.3 Statistical analysis**

6 Descriptive analyses of the study group were given as numbers and percentages. SPSS
7 Statistics 22.0 (IBM SPSS, 2013, Armonk, NY) was used for statistical analysis.
8 Comparisons between recurrent and non-recurrent cases were made using the Chi-square
9 and Fisher's Exact test, and the comparison of the Group 1 and Group 2 with the Mc
10 Nemar test. **The mean and standart deviation values were compared with Student t test.**
11 ~~The median and standard deviation values of nonparametric continuous variables were~~
12 ~~compared with the Mann-Whitney U test.~~ In all analyses, the statistical significance level
13 was taken as $p = 0.05$. The effects of recurrence, PD-L1 expression, MMR status, and
14 loss of PMS2 on survival in patients were examined using log-rank test. Survival rates
15 were calculated using the Kaplan-Meier survival analysis.

16 **3. Results**

17 Of the 74 GBM patients, 29 (39.2%) were female and 45 (60.8%) were male with a
18 median age of 58.4 (range, 4-85). However, the mean age of diagnosis in recurrent cases
19 was 52.1 (4-75) and 62.1 (42-85) in non-recurrent cases and was significantly lower in
20 recurrent GBM cases ($p = 0.007$). Twenty-seven cases (36.5%) recurred. Sixty-two cases
21 (83.8%) died and 12 cases (25.5%) are still alive. The mean recurrence time was 8.2
22 months (0.8-39.7). Median survival was 9.9 months (6.1-13.8) in all cases from the time
23 of diagnosis, and 12.5 months in recurrent cases; 6.0 months for non-recurrent ones. The
24 clinicopathological characteristics of the patients are given in Table 1.

1 On immunohistochemical study, PD-L1 expression was observed in 36 (48.6%) of the 74
2 cases. Expression was detected in 12 (44.5%) of the recurrent cases and 24 (51.0%) of
3 the non-recurrent cases. Median survival was 7.1 months in patients with PD-L1 (+) and
4 10.0 months in patients with PD-L1 (-). The effect of PD-L1 expression on survival was
5 not significant ($p = 0.300$), shown in Figure 3. Thirteen (17.6%) patients exhibited loss
6 of expression for at least one MMR protein and they were considered to MSI. Nine of the
7 cases with MSI were recurrent (33.3%) and 4 were non-recurrent GBM (8.5%). The
8 details of the immunohistochemical expression of MMR proteins are given in Table 2-3.
9 Median survival for tumors with MSI was 7.0 months, and 10.0 months for those with
10 MSS. However, MMR status did not have a significant effect on survival ($p = 0.953$).
11 Among the MMR proteins, loss of PMS2 was noted in 9 recurrent cases (33.3%) and in
12 4 non-recurrent cases (8.5%). Median survival was 7.0 months in patients with PMS2 loss
13 and 10.0 months in patients without loss. However, there was no significant effect of
14 PMS2 loss on survival ($p = 0.953$). Loss of PMS2 was found to be significantly effective
15 in relapsed cases ($p = 0.003$). The median recurrence time was 10.1 months in those with
16 PMS2 loss, and 39.7 months in those without. PD-1 expression was observed in 6 cases
17 (8.1%). It was observed in 3 recurrent cases (11.1%) and 3 non-recurrent cases (6.3%).
18 The expressions of PD-L1, PD-1 and presence of MSI are given in Table 4.

19 Clinical features of Group 1 and 2 (recurrent) cases: 11 (40.7%) were female and 16
20 (59.3%) were male with the median age was 52.1 (range, 4-75). All of them died. The
21 number of cases with PD-L1 (+) in Group 1 was 8 (29.6%). MSI was detected in 6 of 27
22 cases (22.2%). The number of the cases with PD-1 (+) in Group 1 was 1 (3.7%). Loss of
23 PMS2 was observed in 5 cases (18.5%). The number of cases with PD-L1 (+) in Group 2
24 was 8 (29.6%), of which 4 were the same cases in Group 1. MSI was detected in 6 of 27
25 cases (22.2%). Three of these cases were the same as Group 1. The number of cases with

1 PD-1 (+) was 2 (7.4%), both cases were different from Group 1. Six cases had PMS2 loss
2 (22.2%). Two cases were the same as Group 1 and PMS2 loss was observed in 9 cases
3 totally. Clinical characteristics of Group 3 cases: 18 (38.3%) were women and 29 (61.7%)
4 were men with the median age was 62.1 (42-85). Thirty five cases (74.5%) died and 12
5 cases (25.5%) are still alive.

6 The relationship between MMR status and PD-L1 expression in recurrent and non-
7 recurrent cases was shown in Figure 4 ($p = 0.448$ and $p = 0.348$, respectively, **Chi-square**
8 **Fisher's Exact test**). There was no significant difference between PMS2 and PD-L1
9 expression in patients with or without recurrence ($p = 1.000$ and $p = 0.348$, respectively,
10 **Chi-square Fisher's Exact test**). There was no significant difference between PD-L1
11 expression and survival in patients with or without recurrence ($p = 0.136$, **Log Rank**
12 **(Mantel Cox test)**). There was no significant difference between MMR status and survival
13 in patients with or without recurrence ($p = 0.133$, **Log Rank (Mantel Cox test)**). There
14 was no significant difference between loss of PMS2 and survival in patients with or
15 without recurrence ($p = 0.133$, **Log Rank (Mantel Cox test)**).

16 **4. Discussion**

17 The ability of GBM to cause local and systemic immunosuppression limits the innate
18 defense and adaptive immunotherapy effect against the tumor, and thus prevents the
19 development of new therapies. The process of immunosuppression is not only related to
20 abnormal PD-L1 expression in GBM cells, but also to the tumor microenvironment.

21 In one of the most recent studies to elucidate immunotherapy resistance mechanisms in
22 GBM, it was seen that low PD-L1 expression, low tumor mutation burden and T
23 lymphocytes, which are largely depleted in the tumor, are indicators of decreased anti-
24 tumor immunity [21].

1 However, PD-L1 expression levels are observed in a highly variable range in GBMs.
2 Although it was seen between 61% and 88% in two studies with large patient groups, the
3 median percentage of PD-L1 expression on tumor cells in the study of Nudom et al. was
4 2.7% [13-15]. Berghof et al. said that the rate of PD-L1-positive cases in GBM was quite
5 higher (72% in recurrent; 88% in newly diagnosed GBM) than melanoma cases (30%)
6 and non-small cell lung cancer cases (25-36%) [13]. In recurrent GBM cases with higher
7 mutations and where immunotherapy resistance is expected less, we also found PD-L1
8 expression lower than those which are non-recurrent (44.5%, and 51.0% respectively),
9 similar to the studies of Berghof, Heynckes and Ndom [13,14,18].

10 While low PD-L1 expression levels are associated with treatment resistance, some studies
11 have shown that high PD-L1 levels are associated with shorter overall survival in glioma
12 patients [12,17,29]. On the other hand, in several other studies no significant relationship
13 was found between PD-L1 expression and survival [12,13,16,30]. In our study, no
14 significant difference was found between PD-L1 expression level and survival.

15 In tumors with deficient MMR, 10 to 100 times more somatic mutations were found
16 compared to those which are proficient [21,31]. Microsatellite instability is not high in
17 GBM. Patients with MSI are generally young and have colorectal cancer at the same time
18 [32]. In a study conducted with 30 different tumors, it was stated that the neontigen burden
19 in GBM was in the lower third section [21,33]. In GBM, mutations in MMR genes are
20 thought to be associated with resistance to therapy and thus tumor recurrence [8]. Martine
21 et al. observed that the presence of MSI was at a significantly higher rate in patients with
22 recurrent GBM and stated that this may be associated with malignant progression [34].
23 We have also found the rate of MSI significantly higher in recurrent patients than the non-
24 recurrent ones (33.3%, and 8.5% respectively).

1 GBM specimens containing MSH6 mutations have been described as hypermutator
2 phenotypes [35,36]. Shinsato et al. found reduced levels of MLH1 and PMS2 related to
3 therapy resistance and recurrence [37]. We have also found a significant elevation in
4 PMS2 loss in all groups (Table 2-3). PMS2 loss, which was observed more clearly in
5 recurrent cases, suggested that this change might be a marker for malignant progression.
6 However, we could not find a significant relationship between the loss of PMS2 neither
7 with survival nor PD-L1 expression.

8 In one of the studies on the role of the status of MSI in predicting immunotherapy
9 response, it was observed that colorectal cancers with MMR deficiency had a high
10 response to PD-1 inhibitor therapy [30]. In another study, the research was expanded and
11 the effectiveness of PD-1 blockade was evaluated in 12 different tumor types with MMR
12 deficiency at an advanced stage and it was seen that 21% of the patients had a complete
13 response and 53% of the patients had an an objective radiological response [38]. In high-
14 grade urothelial carcinomas, it was shown that MMR deficiency (loss of MSH2 and
15 MSH6) is associated with increased PD-L1 expression [8,39]. PD-1 and PD-L1
16 expressions were found high in colorectal and endometrial cancers with microsatellite
17 instability (MSI) [8,40].

18 In conclusion, MMR status is suggested as a marker for response to PD-1 / PD-L1
19 blockade in other cancer types. However, we found that PD-L1 expression was low and
20 MSI rate was higher in recurrent GBM than non-recurrent cases and we could not find a
21 significant relationship between these two entities. The presence of higher MSI in the
22 patients with recurrent GBM in this study indicates the importance of these proteins as a
23 predictive markers. However, low PD-L1 expression levels suggest that this antibody
24 may not be a good predictive marker for determining the group of patients who will
25 receive immunotherapy.

1 In recent years, an increasing number of clinical trials are available to try different
2 combinations in GBM treatment. We also think that different combinations with other
3 immune checkpoint proteins can be tried in GBM patients to determine both prognostic
4 and therapeutic efficacy.

5 The limitation of our study was that the clinical data about treatment modalities are
6 incomplete and, therefore, were not included in the article.

7 **Acknowledgement and/or disclaimers**

8 This project (Project no: TSB-2019-7796) was supported by the Karadeniz Technical
9 University Scientific Research Projects Support Fund. Approval of Karadeniz Technical
10 University Faculty of Medicine Ethics Committee was obtained before starting the study
11 (26.07.2018- Protocol no: 2018/167).

12 **References**

- 13 1. Ostrom QT, Gittleman H, Liao P, Vecchione-Koval T, Wolinsky Y et al.
14 CBTRUS Statistical Report: Primary brain and other central nervous system
15 tumors diagnosed in the United States in 2010-2014. *Neuro-Oncology* 2017;
16 19(5):1-88. doi: 10.1093/neuonc/nox158
- 17 2. Krex D, Klink B, Hartmann C, von Deimling A, Pietsch T et al. Long-term
18 survival with glioblastoma multiforme. *Brain* 2007; 130(10):2596-2606. doi:
19 10.1093/brain/awm204
- 20 3. Curran WJ Jr, Scott CB, Horton J, Nelson JS, Weinstein AS et al. Recursive
21 partitioning analysis of prognostic factors in three Radiation Therapy Oncology
22 Group malignant glioma trials. *Journal of the National Cancer Institute* 1993;
23 85(9):704-710. doi: 10.1186/1471-2407-9-450

- 1 4. Zheng T, Cantor KP, Zhang Y, Chiu BC, Lynch CF. Risk of brain glioma not
2 associated with cigarette smoking or use of other tobacco products in Iowa.
3 *Cancer Epidemiology, Biomarkers & Prevention* 2001; 10(4):413-414.
- 4 5. Huncharek M, Kupelnick B, Wheeler L. Dietary cured meat and the risk of adult
5 glioma: a meta-analysis of nine observational studies. *Journal of Environmental*
6 *Pathology, Toxicology and Oncology* 2003; 22(2):129-137. doi:
7 10.1615/JEnvPathToxOncol.v22.i2.60
- 8 6. Kleihues P, Schäuble B, zur Hausen A, Estève J, Ohgaki H. Tumors associated
9 with p53 germline mutations: a synopsis of 91 families. *The American Journal of*
10 *Pathology* 1997; 150(1):1-13.
- 11 7. Reardon DA, Gokhale PC, Klein SR, Ligon KL, Rodig SJ et al. Glioblastoma
12 eradication following immune checkpoint blockade in an orthotopic,
13 immunocompetent model. *Cancer Immunology Research* 2016; 4 (2): 124-135.
14 doi: 10.1158/2326-6066.CIR-15-0151
- 15 8. Kurz SC, Wen PY. Quo Vadis-Do Immunotherapies Have a Role in
16 Glioblastoma? *Current Treatment Options in Neurology* 2018; 20(5):14. doi:
17 10.1007/s11940-018-0499-0
- 18 9. Wing K, Onishi Y, Prieto-Martin P, Yamaguchi T, Miyara M et al. CTLA-4
19 control over Foxp3+ regulatory T cell function. *Science* 2008; 322(5899):271-
20 275. doi: 10.1126/science.1160062
- 21 10. Bloch O, Crane CA, Kaur R, Safaee M, Rutkowski MJ et al. Gliomas promote
22 immunosuppression through induction of B7-H1 expression in tumor-associated
23 macrophages. *Clinical Cancer Research* 2013;19(12):3165-3175. doi:
24 10.1158/1078-0432.CCR-12-3314

- 1 11. Chen PL, Roh W, Reuben A, Cooper ZA, Spencer CN et al. Analysis of Immune
2 Signatures in Longitudinal Tumor Samples Yields Insight into Biomarkers of
3 Response and Mechanisms of Resistance to Immune Checkpoint Blockade.
4 *Cancer Discovery* 2015; 6(8):827-37. doi: 10.1158/2159-8290.CD-15-1545
- 5 12. Chen RQ, Liu F, Qiu XY, Chen XQ. The Prognostic and Therapeutic Value of
6 PD-L1 in Glioma. *Frontiers in Pharmacology* 2019; 9:1503. doi:
7 10.3389/fphar.2018.01503
- 8 13. Berghoff AS, Kiesel B, Widhalm G, Rajky O, Ricken G et al. Programmed death
9 ligand 1 expression and tumor-infiltrating lymphocytes in glioblastoma. *Neuro-*
10 *Oncology* 2015; 17(8):1064-75. doi: 10.1093/neuonc/nou307
- 11 14. Nduom EK, Wei J, Yaghi NK, Huang N, Kong LY et al. Heimerger AB: PD-L1
12 expression and prognostic impact in glioblastoma. *Neuro-Oncology* 2016;
13 18(2):195-205. doi: 10.1093/neuonc/nov172
- 14 15. Preusser M, Berghoff AS, Wick W, Weller M. Clinical Neuropathology mini-
15 review 6-2015: PD-L1: emerging biomarker in glioblastoma? *Clinical*
16 *Neuropathology* 2015; 34(6):313-321. doi: 10.5414/NP300922
- 17 16. Zeng J, Zhang XK, Chen HD, Zhong ZH, Wu QL et al. Expression of programmed
18 cell death-ligand 1 and its correlation with clinical outcomes in gliomas.
19 *Oncotarget* 2016; 7(8):8944-8955. doi: 10.18632/oncotarget.6884
- 20 17. Xue S, Song G, Yu J. The prognostic significance of PD-L1 expression in patients
21 with glioma: A meta-analysis. *Scientific Reports* 2017; 7(1):4231. doi:
22 10.1038/s41598-017-04023-x
- 23 18. Heynckes S, Gaebelein A, Haaker G, Grauvogel J, Franco P et al. Expression
24 differences of programmed death ligand 1 in de-novo and recurrent glioblastoma

- 1 multiforme. Oncotarget 2017; 8(43): 74170–74177. doi:
2 10.18632/oncotarget.18819
- 3 19. Bouffet E, Larouche V, Campbell BB, Merico D, de Borja R, et al. Immune
4 Checkpoint Inhibition for Hypermutant Glioblastoma Multiforme Resulting From
5 Germline Biallelic Mismatch Repair Deficiency. *Jornal of Clinical Oncology*
6 2016; 34(19):2206-2211. doi: 10.1200/JCO.2016.66.6552
- 7 20. Llosa NJ, Cruise M, Tam A, Wicks EC, Hechenbleikner EM et al. The vigorous
8 immune microenvironment of microsatellite instable colon cancer is balanced by
9 multiple counter-inhibitory checkpoints. *Cancer Discovery* 2015; 5(1):43-51. doi:
10 10.1158/2159-8290
- 11 21. Adhikaree J, Moreno-Vicente J, Kaur AP, Jackson AM, Patel PM. Resistance
12 Mechanisms and Barriers to Successful Immunotherapy for Treating
13 Glioblastoma. *Cells* 2020; 9(2):263. doi: 10.3390/cells9020263
- 14 22. Zou W, Wolchok JD, Chen L. PD-L1 (B7-H1) and PD-1 pathway blockade for
15 cancer therapy: Mechanisms, response biomarkers, and combinations. *Science*
16 Translational Medicine 2016; 8(328):328rv4. doi: 10.1126/scitranslmed.aad7118
- 17 23. Zeng J, See AP, Phallen J, Jackson CM, Belcaid Z et al. Anti-PD-1 blockade and
18 stereotactic radiation produce long-term survival in mice with intracranial
19 gliomas. *International Journal of Radiation Oncology Biology, Physics* 2013; 86
20 (2): 343-349. doi: 10.1016/j.ijrobp.2012.12.025
- 21 24. Roth P, Valavanis A, Weller M. Long-term control and partial remission after
22 initial pseudoprogression of glioblastoma by anti-PD-1 treatment with nivolumab.
23 *Neuro-Oncology* 2017; 19(3):454-456. doi: 10.1093/neuonc/now265

- 1 25. Simonelli M, Di Tommaso L, Baretta M, Santoro A. Pathological characterization
2 of nivolumab-related liver injury in a patient with glioblastoma. *Immunotherapy*
3 2016; 8(12):1363-1369. doi: 10.2217/imt-2016-0057
- 4 26. Tihan T, Barletta J, Parney I, Lamborn K, Sneed PK et al. Prognostic value of
5 detecting recurrent glioblastoma multiforme in surgical specimens from patients
6 after radiotherapy: should pathology evaluation alter treatment decisions?. *Human*
7 *Pathology* 2006; 37(3):272-282. doi: 10.1016/j.humpath.2005.11.010
- 8 27. Felsberg J, Thon N, Eigenbrod S, Hentschel B, Sabel MC et al. Promoter
9 methylation and expression of MGMT and the DNA mismatch repair genes
10 MLH1, MSH2, MSH6 and PMS2 in paired primary and recurrent glioblastomas.
11 *International Journal of Cancer* 2011; 129(3):659-670. doi: 10.1002/ijc.26083
- 12 28. Collage of American pathologist (2018). Template for Reporting Results of DNA
13 Mismatch Repair Testing in Patients Being Considered for Checkpoint Inhibitor
14 Immunotherapy (online). Website: [https://documents.cap.org/protocols/cp-](https://documents.cap.org/protocols/cp-general-dnamismatchrepair-18biomarker-1001.pdf)
15 [general-dnamismatchrepair-18biomarker-1001.pdf](https://documents.cap.org/protocols/cp-general-dnamismatchrepair-18biomarker-1001.pdf) (accessed 09.10.2020)
- 16 29. Wang Z, Zhang C, Liu X, Wang Z, Sun L et al. Molecular and clinical
17 characterization of PD-L1 expression at transcriptional level via 976 samples of
18 brain glioma. *Oncoimmunology* 2016; 5(11):e1196310. doi:
19 10.1080/2162402X.2016.1196310
- 20 30. Miyazaki T, Ishikawa E, Matsuda M, Akutsu H, Osuka S et al. Assessment of PD-
21 1 positive cells on initial and secondary resected tumor specimens of newly
22 diagnosed glioblastoma and its implications on patient outcome. *Journal of Neuro-*
23 *Oncology* 2017; 133(2):277-285. doi: 10.1007/s11060-017-2451-7

- 1 31. Le DT, Uram JN, Wang H, Bartlett BR, Kemberling H et al. PD-1 Blockade in
2 Tumors with Mismatch-Repair Deficiency. *The New England Journal of*
3 *Medicine* 2015; 372(26):2509-2520. doi: 10.1056/NEJMoa1500596
- 4 32. Leung SY, Chan TL, Chung LP, Chan AS, Fan YW et al. Microsatellite Instability
5 and Mutation of DNA Mismatch Repair Genes in Gliomas. *The American Journal*
6 *of Pathology* 1998; 153(4): 1181–1188. doi: 10.1016/S0002-9440(10)65662-3
- 7 33. Schumacher TN, Schreiber RD: Neoantigens in cancer immunotherapy. *Science*
8 2003; 348(6230):69-74. doi: 10.1126/science.aaa4971
- 9 34. Martinez R, Schackert HK, Plaschke J, Baretton G, Appelt H et al. Molecular
10 Mechanisms Associated with Chromosomal and Microsatellite Instability in
11 Sporadic Glioblastoma multiforme. *Oncology* 2004; 66:395–403. doi:
12 10.1159/000079488
- 13 35. Hodges TR, Ott M, Xiu J, Gatalica Z, Swensen J et al. Mutational burden, immune
14 checkpoint expression, and mismatch repair in glioma: implications for immune
15 checkpoint immunotherapy. *Neuro-Oncology* 2017; 19(8):1047-1057. doi:
16 10.1093/neuonc/nox026
- 17 36. Cahill DP, Levine KK, Betensky RA, Codd PJ, Romany CA et al. Loss of the
18 mismatch repair protein MSH6 in human glioblastomas is associated with tumor
19 progression during temozolomide treatment. *Clinical Cancer Research* 2007;
20 13(7): 2038–2045. doi: 10.1158/1078-0432.CCR-06-2149
- 21 37. Shinsato Y, Furukawa T, Yunoue S, Yonezawa H, Minami K et al. Reduction of
22 MLH1 and PMS2 confers temozolomide resistance and is associated with
23 recurrence of glioblastoma. *Oncotarget* 2013; 4(12): 2261–2270. doi:
24 10.18632/oncotarget.1302

- 1 38. Le DT, Durham JN, Smith KN, Wang H, Bartlett BR et al. Mismatch-repair
2 deficiency predicts response of solid tumors to PD-1 blockade. *Science* 2017;
3 357(6349): 409–413. doi: 10.1126/science.aan6733
- 4 39. Castro MP, Goldstein N. Mismatch repair deficiency associated with complete
5 remission to combination programmed cell death ligand immune therapy in a
6 patient with sporadic urothelial carcinoma: immunotheranostic considerations.
7 *Journal for Immunotherapy of Cancer* 2015; 3:58. doi: 10.1186/s40425-015-0104-
8 y
- 9 40. Yamashita H, Nakayama K, Ishikawa M, Nakamura K, Ishibashi T et al.
10 Microsatellite instability is a biomarker for immune checkpoint inhibitors in
11 endometrial cancer. *Oncotarget* 2018; 9(5): 5652–5664. doi:
12 10.18632/oncotarget.23790

13
14
15
16
17
18
19
20
21
22
23
24
25
26

1 **Table 1:** The clinicopathological characteristics of the recurrent and non-recurrent cases

Clinical parameters	Recurrent cases (n=27)	Non-recurrent cases (n=47)	Total (n=74)
Gender (Female/Male) n (%)	11 (40.7) / 16 (59.3)	18 (38.3) / 29 (61.7)	29 (39.2) / 45 (60.8)
Mean age (\pm SD)	52.1 (\pm 15.349)	62.1 (\pm 10.693)	58.4 (\pm 13.392)
Mean recurrence time (month) (95% CI)	8.2 (0.8-39.7)	-	-
Median survival time (month) (95% CI)	12.5 (7.4-17.5)	6.0 (3.7-8.2)	9.9 (6.1-13.8)
Death n (%)	27 (100)	35 (74.5)	62 (83.8)

2 Student t test: p=0,02

3 Median survival time: Kaplan-Meier

4

5

6

7

8

9

10

11

12

13

14

15

16

1 **Table 2:** Loss of MMR proteins in the recurrent cases (n=27, p < 0.05)

MMR proteins	Resection material	n	%	p
MLH1	1st resection	1	3.7	-
	2nd resection	0	0	
MSH2	1st resection	0	0	-
	2nd resection	0	0	
MSH6	1st resection	0	0	-
	2nd resection	2	7.4	
PMS2	1st resection	5	18.5	1.000
	2nd resection	6	22.2	

2 MMR: Mismatch repair

3 **McNemar Exact Test**

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

1 **Table 3:** Loss of MMR proteins in all cases (n=74, p < 0.05)

MMR proteins	Cases	n	%	p
MLH1	Recurrent	1	3.7	0.635
	Non-recurrent	1	2.1	
MSH2	Recurrent	0	0	-
	Non-recurrent	0	0	
MSH6	Recurrent	2	7.4	0.530
	Non-recurrent	2	4.3	
PMS2	Recurrent	9	33.3	0.011*
	Non-recurrent	4	8.5	

2 MMR: Mismatch repair

3 *Statistically significant

4

5 **Chi-Square Fisher's Exact test**

6

7

8

9

10

11

12

13

14

15

16

17

1 **Table 4:** The expressions of PD-L1, PD-1 and presence of MSI in the recurrent and
2 non-recurrent cases (n=74, p < 0.05)

Protein expression	Cases	n	%	p
PD-L1	Recurrent	12	44.5	0.060
	Non-recurrent	24	51.0	
PD-1	Recurrent	3	11.1	0.536
	Non-recurrent	3	6.4	
MSI	Recurrent	9	33.3	0.011*
	Non-recurrent	4	8.5	

3 MSI: Microsatellite instability

4 *Statistically significant

5

6 **Chi-Square Fisher's Exact test**

7

8

9

10

11

12

13

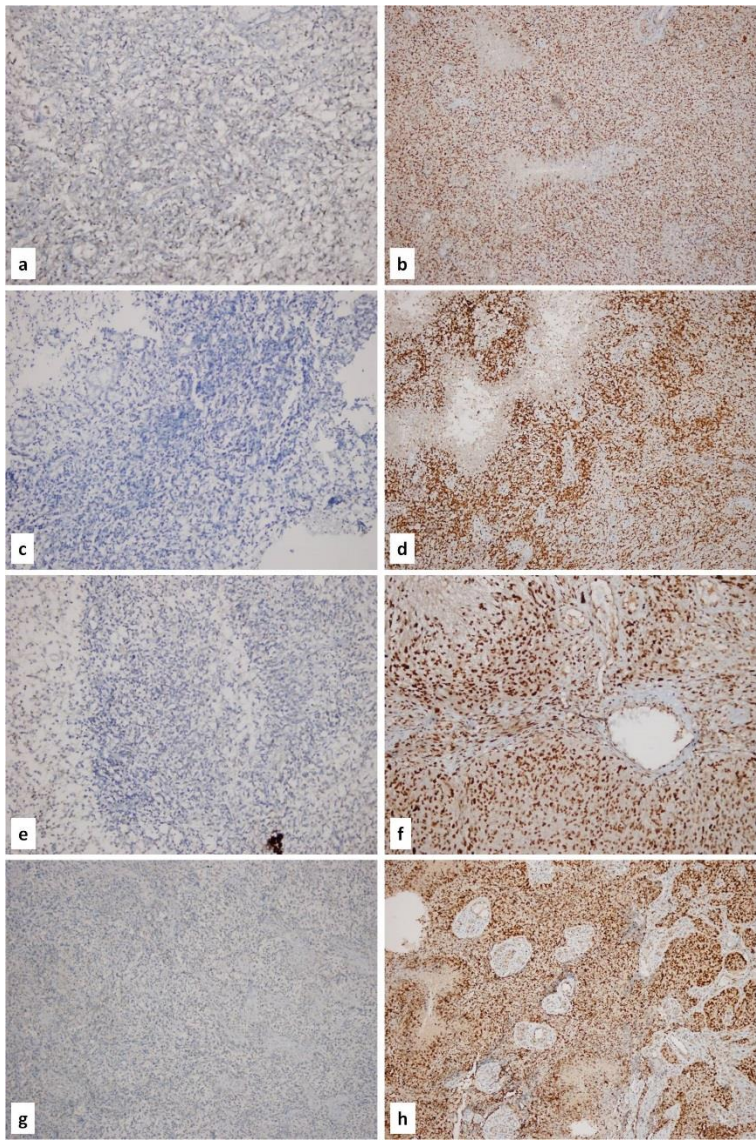
14

15

16

17

1 **Figures**



2

3 Figure 1: Expressions of MMR proteins.

4

5

6

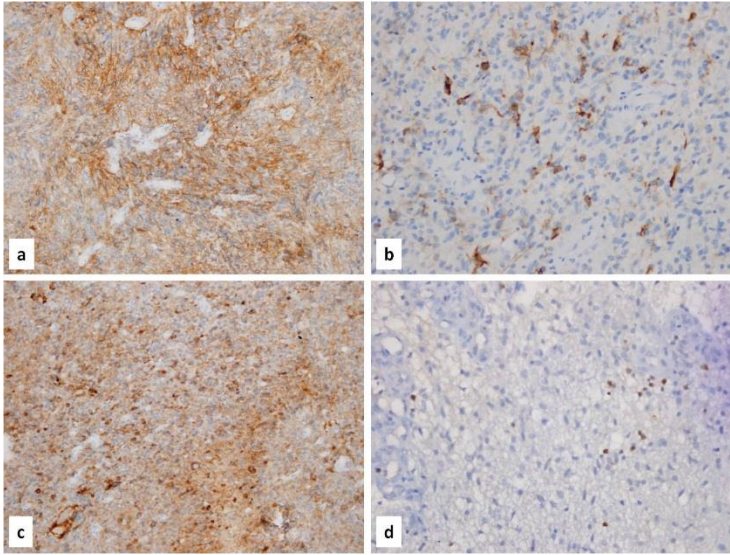
7

8

9

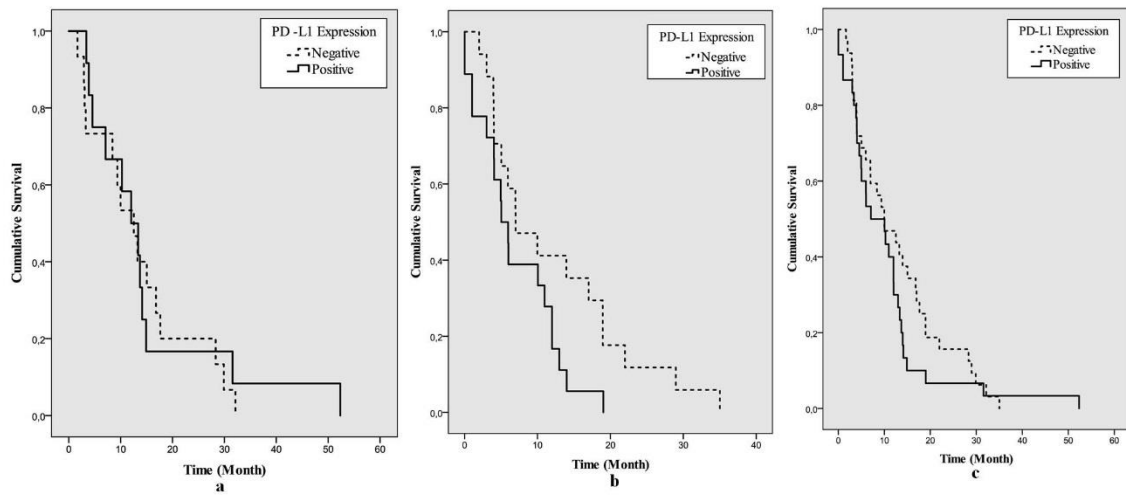
10

11



1
2
3
4
5
6
7
8
9
10
11
12
13

Figure 2: The staining patterns of PD-L1 and PD-1.



1

2

3 Figure 3: The effect of PD-L1 expression on survival.

4

5

6

7

8

9

10

11

12

13

14

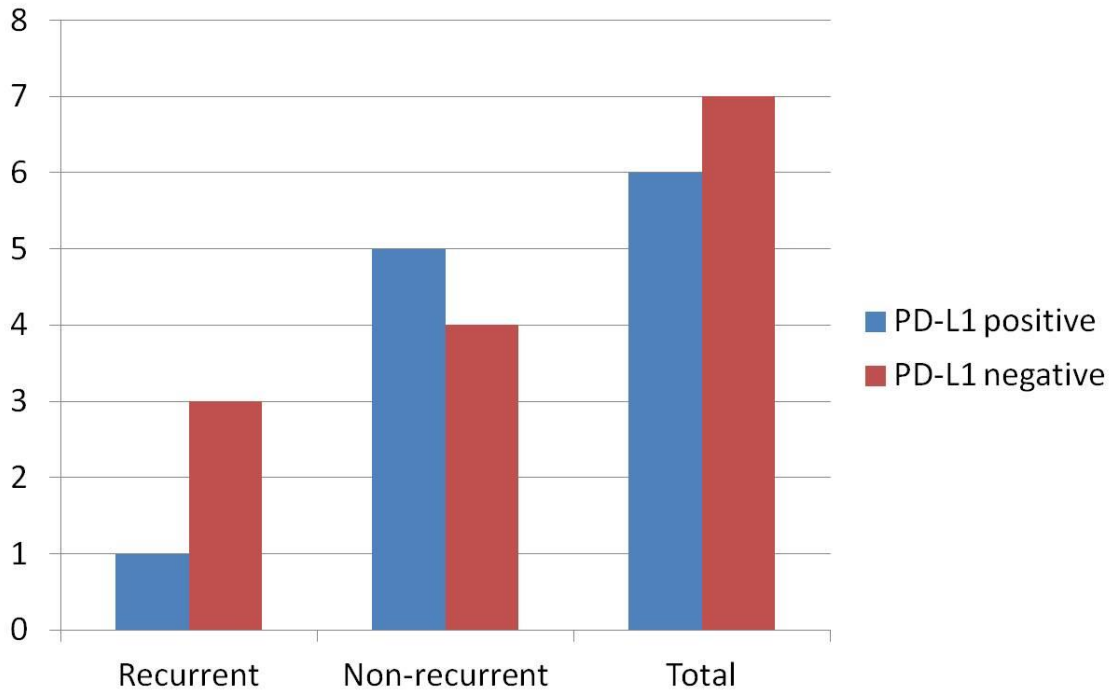
15

16

17

18

19



1
2
3
4
5
6
7
8
9
10
11
12
13
14
15

Figure 4: The relationship between MMR status and PD-L1 expression in recurrent and non-recurrent GBM.