Abstract

**Background/aim:** In this study, we aimed to evaluate the initial hematological findings analyzed on admission in confirmed COVID-19 patients who were transferred to the intensive care unit (ICU), to predict possible hematological indices.

**Materials and methods:** Initial neutrophil to lymphocyte ratio (NLR), platelet to lymphocyte ratio (PLR), monocyte to lymphocyte ratio (MLR), red cell distribution width to platelet ratio (RPR), mean platelet volume to platelet ratio, and lymphocyte multiplied by platelet count (LYM x PLT), of 695 patients with laboratory-confirmed COVID-19 were investigated and compared between the mild/moderate and severe groups.

**Results:** The proportion of COVID-19 cases admitted to ICU was 3.9%. The median age of patients admitted to ICU was significantly higher than those who were not; (68.5 (interquartile range (IQR); 21.5) years vs. 41.0 (IQR; 15.7) years; p <0.001). Severe cases had higher NLR (6.6 vs 2.4; P <0.001), and MLR (0.40 vs 0.28; P=0.004) and lower PLR (180.0 vs 129.0; P <0.001) compared to that of mild or moderate patients. Among all of the parameters, the ROC curve of NLR gave us the best ability to distinguish serious patients at an early stage (AUC = 0.819, 95% confidence interval 0.729–0.910; p<0.001).

**Conclusion:** These data showed that age, initial NLR, PLR, and LYM x PLT were associated with the severity of COVID-19 disease and patients’ need for the ICU. Therefore, initial hemogram parameters may be essential to predict the prognosis of COVID-19 patients.

**Key words:** Intensive care unit, COVID-19, neutrophils, leukocytes, platelets
**Introduction**

In December 2019, a novel coronavirus, severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), was identified in Wuhan, China; and quickly spread around the World [1]. The World Health Organization named this infection, as COVID-19 and announced as a pandemic [2]. In our country, the first PCR positive COVID-19 case was detected on March 11th, 2020 [3].

The clinical spectrum of COVID-19 infection can vary from asymptomatic to severe disease (e.g., acute respiratory distress syndrome [ARDS], acute cardiac injury, and acute kidney injury) [1, 4]. Up to 32% of all positive patients require intensive care unit (ICU) admission, and death can occur [4, 5]. Therefore, early identification of patients with severe disease risk, and those potentially developing a life-threatening condition is important.

Increasing evidence supports the role of a dysfunctional immune response in the airway damage and the progression of various viral pneumonia, including COVID-19 [6, 7]. Hematological indices, such as neutrophil to lymphocyte ratio (NLR), platelet to lymphocyte (PLR) ratio, monocyte to lymphocyte ratio (MLR), lymphocyte count multiplied by platelet count (LYM x PLT), red cell distribution width to platelet ratio (RPR), are indicators of the systemic inflammatory response and has been extensively investigated in various diseases [8-11]. Abnormal blood count results were detected in patients with COVID-19 [6, 12-14], which are considered to be potential predictors of the outcomes [15, 16].

In this study, we aimed to identify hematological indices of confirmed COVID-19 patients that were analyzed on admission and their relationship with the disease severity in order to be used for predicting the disease progression.
Materials and methods

The study was approved by the ethics committee of SBU Bursa Yüksek İhtisas Training, and Research Hospital (No. 2020/05-02); a pandemic hospital with 1430 beds served as the setting for the present retrospective cohort study. The study was conducted in accordance with the principles outlined in the Declaration of Helsinki. Combined nasopharyngeal and oropharyngeal swabs were taken from 1100 patients who met the suspected case definitions for COVID-19. Real-time quantitative amplification of SARS CoV RNA was performed with SARS-CoV-2 (2019-nCoV) RT-qPCR Detection Kit (Bio-Speedy®, Bioeksen R&D Technologies, Istanbul, Turkey) with a thermal cycler device (CFX96TM Real-Time System, BIO-RAD, Singapore) according to the manufacturer's instructions. Six-hundred and ninety-five confirmed SARS-CoV-2 patients admitted to our hospital between March 27th, and April 30th, 2020, were included in the study. Individuals, younger than 18 years old, required aggressive treatment within 24 hours of admission, and absence of initial complete blood count (CBC) test results were excluded from the study.

Twenty-seven patients who fulfilled one of the following criteria; dyspnea and respiratory distress; respiratory rate> 30/min, PaO2 / FiO2 <300; SPO2 <90 despite 5 L/min oxygen treatment, PaO2 <70, hypotension (systolic blood pressure <90 mmHg and/or more than 40 mmHg decrease and mean arterial pressure <65 mmHg, tachycardia >100/min, acute kidney injury, development of acute organ dysfunction, and patients with immunosuppression, acute bleeding diathesis, troponin increase, arrhythmia, lactate> 2 mmol, capillary return disorder, and cutis marmoratus were transferred to ICU.
The initial triage protocol of COVID-19 patients involves drawing blood samples upon admission. CBCs were analyzed within 1 hour after venipuncture using an automatic blood counter (Mindray BC-5800; Mindray Biomedical Electronics Co., Ltd., Shenzhen, People's Republic of China). Mean platelet volume divided by platelet count (MPV to PLT) and lymphocyte multiplied by platelet (LYM x PLT) were recorded. NLR, PLR, and MLR were calculated by dividing the number of neutrophils, platelet, and monocyte count by the lymphocyte count. RDW to platelet ratio (RPR) was calculated using the formula (RDW \times 100 / PLT (10^9/L)) [17].

**Statistical Analysis**

Statistical analysis was performed using SPSS (Statistical Package for Social Sciences) for Windows (SPSS Inc, Chicago, IL), and P values of < 0.05 were considered significant. Descriptive statistics will be presented as frequency distribution and percentage. The Kolmogorov-Smirnov test was used to identify the distribution of variables. Statistically significant differences between the variables were established using the Mann–Whitney U test. A binary logistic regression analysis was performed to determine the influence of age and all other significant factors.

**Results**

The proportion of COVID-19 cases admitted to ICU was 3.9%. Signs and symptoms of severe cases on admission included self-reported fever (52%), cough (44%), shortness of breath (52%), sore throat (19%), myalgia/arthritis (19%), fatigue (15%), chest pain/discomfort (11%), nasal symptoms (11%), headache (11%), nausea/vomiting (11%) and diarrhea (4%).
During the study period, the median age of patients who were transferred to ICU was significantly higher than those who were not transferred (68.5 (interquartile range (IQR); 21.5) years vs. 41.0 (IQR; 15.7) years; p <0.001, Table 1).

When comparing the initial hemoglobin concentration (12.9 (4.0) vs. 13.8 (2.3) g / dL; P <0.001) and lymphocyte count (12.7% (12.6) vs. 26.6% (15.7)) both values were found to be lower than that of non-severe cases (Table 1).

It is observed that severe cases had higher NLR (6.6 vs. 2.4; P <0.001), and MLR (0.40 vs. 0.28; P=0.004) and lower PLR (180.0 vs. 129.0; P <0.001) compared to mild or moderate patients (Table 1). The ROC curve of NLR gave us the best prediction opportunity for distinguishing patients with severe disease from an earlier stage (AUC = 0.819, 95% confidence interval 0.729–0.910; p<0.001, Table 2).

Potential risk factors, including age, NLR, PLR, LYM x PLT, and RDW were investigated using binary logistic regression analysis. Age was found to be the only significant (b=0.070, SE=0.015, p<0.001) positive predictor for ICU requirement, with the OR 1.073 (95% CI, 1.042 to 1.104) (Table 3).

**Discussion**

On admission, the COVID-19 patients whom needed to be taken to the ICU were older than the other, as reported before [1, 4]. In the ICU-patient group, the initial absolute neutrophil count was higher, and the lymphocyte count was lower among patients who were transferred to the ICU compared to mild cases who were not. This is in accordance with the previous publications that reported the increase in leukocyte count and a decrease in lymphocyte count as an indicator for clinical deterioration in COVID patients [18, 19].
Neutrophils are the most important cellular defense line against infections, which first responds to viral invasion, and limits the viral replication and spread. However, neutrophils have also been reported to mediate deleterious effects on the host during viral infection [20].

The immune response to a viral infection is mainly based on lymphocytes. It has been assumed that a significant decrease in lymphocyte count may be due to increased lymphocyte consumption, destruction of lymphatic tissues, and cytokine-induced T-cell apoptosis in patients with COVID-19 [15, 21]. Lymphopenia, as a sign of the severe disease, is not specific to COVID-19. It has also been seen in other viral causes of pneumonia, such as influenza [22, 23].

Notably, we observed that COVID-19 patients who were transferred to the ICU had lower lymphocyte count and higher NLR and PLR. In consistent with our study, Yang et al. suggested NLR ≥3.3 as an indicator with clinical symptoms to change disease status from mild to severe disease [24]. NLR is reported to be used as an early indicator for the severe disease, similar to previous findings from a different patient population [24, 25].

In the current study, the median PLR of severe COVID patients was higher compared to that of non-severe cases. The PLR, which reflects both aggregation and inflammation control, is a better marker for the decision of patients for transfer to the ICU than platelet or lymphocyte count alone. Changes in platelet count and activity are closely related to various diseases [26]. It has been shown in several studies that low platelet count is directly related to the severity of the disease in COVID-19 patients [14, 27]. Platelets not only contribute to hemostasis but also participate in the inflammation and
host defense. Decreased platelet production and increased consumption due to diffuse
alveolar damage are thought to cause thrombocytopenia in COVID-19 patients [28, 29].
In accordance with previous studies, the hemoglobin level was found to be lower in
COVID patients who needed to be taken to the ICU when compared to that of milder
COVID patients [30, 31]. RDW is a measure of the change in erythrocyte volume and
has been reported to be a predictor of mortality in some conditions, including infections
[32]. The higher RDW is associated to increased inflammation markers which have
been tightly correlated with the critical disease [33, 34].

Conclusion
According to the results of this study, the variables, including the age of the patients,
NLR, PLR, and LYM x PLT, are related to the severity of COVID-19 disease and may
contribute to decision making for transferring patients to the ICU on admission.
Therefore, initial CBC parameters should be monitored for predicting the prognosis of
COVID-19 patients.

Limitation
This study was a retrospective, single-center study. Prospective and multicenter clinical
studies might be required to avoid a certain degree of deviation and to support the
findings.

Statement on funding sources and conflicts of interest
No financial support and no other potential conflict of interest relevant to this article
were reported.

Acknowledgments
Nothing to declare
References


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**Table 1:** Initial laboratory findings of patients with COVID-19.

<table>
<thead>
<tr>
<th></th>
<th>Group 1 (N=668)</th>
<th>Group 2 (N=27)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, median (IQR)</td>
<td>41.0 (15.7)</td>
<td>69.0 (21.0)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Male/Female (%)</td>
<td>47.3/52.7</td>
<td>55.6/44.4</td>
<td></td>
</tr>
<tr>
<td>WBC count, ×10⁹/L</td>
<td>6.2 (2.95)</td>
<td>8.3 (5.5)</td>
<td>0.008</td>
</tr>
<tr>
<td>Haemoglobin, g/dL</td>
<td>13.8 (2.3)</td>
<td>12.9 (4.0)</td>
<td>0.012</td>
</tr>
<tr>
<td>Hct (%)</td>
<td>40.8 (5.9)</td>
<td>38.5 (10.3)</td>
<td>0.017</td>
</tr>
<tr>
<td>Platelet count, ×10⁹/L</td>
<td>217 (74)</td>
<td>192 (90)</td>
<td>0.081</td>
</tr>
<tr>
<td>NEU (%)</td>
<td>63.8 (16.7)</td>
<td>82.7 (21.5)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>LYM (%)</td>
<td>26.6 (15.7)</td>
<td>12.7 (12.6)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>MONO (%)</td>
<td>7.6 (4.0)</td>
<td>5.9 (5.0)</td>
<td>0.004</td>
</tr>
<tr>
<td>EOS (%)</td>
<td>0.7 (1.3)</td>
<td>0.1 (0.3)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>MCV, μm³</td>
<td>85.7 (5.8)</td>
<td>84.4 (9.0)</td>
<td>0.321</td>
</tr>
<tr>
<td>MCH, pg</td>
<td>29.1 (2.5)</td>
<td>28.2 (4.3)</td>
<td>0.391</td>
</tr>
<tr>
<td>MCHC (g/dL)</td>
<td>33.8 (1.2)</td>
<td>33.3 (1.3)</td>
<td>0.003</td>
</tr>
<tr>
<td>RDW, %</td>
<td>13.1 (1.2)</td>
<td>13.8 (3.1)</td>
<td>0.007</td>
</tr>
<tr>
<td>MPV, fL</td>
<td>9.6 (1.3)</td>
<td>9.7 (1.4)</td>
<td>0.241</td>
</tr>
<tr>
<td>CRP, mg/L</td>
<td>6.2 (13)</td>
<td>73 (123)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>MPV to PLT</td>
<td>0.044 (0.018)</td>
<td>0.053 (0.028)</td>
<td>0.049</td>
</tr>
<tr>
<td>NLR</td>
<td>2.4 (2.0)</td>
<td>6.6 (7.8)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>PLR</td>
<td>129 (70)</td>
<td>180 (156)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>MLR</td>
<td>0.28 (0.2)</td>
<td>0.40 (0.3)</td>
<td>0.004</td>
</tr>
</tbody>
</table>
The patients were divided into two groups: Group 1: Patients not requiring ICU admission; Group 2: ICU admission required. ICU= intensive care unit; IQR= interquartile range; RBC= red blood cell; WBC= white blood cell; Hct= hematocrit; NEU= neutrophil; LYM= lymphocyte; MONO= monocyte; EOS= eosinophils; BASO= basophile; MCV= Mean corpuscular volume; MCH= Mean corpuscular haemoglobin, MCHC= Mean corpuscular hemoglobin concentration, RDW= red cell distribution width; MPV= The mean platelet volume; NLR= neutrophil to lymphocyte ratio; MLR= monocyte to lymphocyte ratio; LYMxPLT= lymphocyte multiplied by platelet; RPR= red cell distribution width to platelet ratio; MPV to PLT= mean platelet volume to platelet count ratio. Data are median (IQR) or n (%).
Table 2: Diagnostic performances of initial hematologic indices for distinguishing admission to ICU.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>AUC (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>NLR</td>
<td>0.819 (0.729-0.910)</td>
</tr>
<tr>
<td>LYM x PLT</td>
<td>0.786 (0.706-0.865)</td>
</tr>
<tr>
<td>PLR</td>
<td>0.746 (0.629-0.862)</td>
</tr>
<tr>
<td>RPR</td>
<td>0.686 (0.584-0.788)</td>
</tr>
<tr>
<td>MLR</td>
<td>0.663 (0.560-0.765)</td>
</tr>
<tr>
<td>MPV to PLT</td>
<td>0.612 (0.492-0.732)</td>
</tr>
</tbody>
</table>

AUC (95% CI) = area under the receiver operating characteristic curve (95% confidence interval); NLR= neutrophil to lymphocyte ratio; LYM= lymphocyte; PLT= platelet; MPV= mean platelet volume; PLR= platelet to lymphocyte ratio; RPR= red cell distribution width to platelet count; MLR= monocyte to lymphocyte ratio; MPV to PLT= mean platelet volume to platelet count ratio.
Table 3: Binary logistic regression analysis predicting the likelihood of intensive care unit admission.

<table>
<thead>
<tr>
<th>Variable</th>
<th>B</th>
<th>S.E.</th>
<th>OR with 95% CI</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age</td>
<td>0.070</td>
<td>0.015</td>
<td>1.073 (1.042-1.104)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Gender</td>
<td>-0.178</td>
<td>0.479</td>
<td>0.837 (0.327-2.141)</td>
<td>0.711</td>
</tr>
<tr>
<td>NLR</td>
<td>0.106</td>
<td>0.073</td>
<td>1.112 (0.964-1.281)</td>
<td>0.145</td>
</tr>
<tr>
<td>LYM x PLT</td>
<td>-0.003</td>
<td>0.002</td>
<td>0.997 (0.994-1.001)</td>
<td>0.209</td>
</tr>
<tr>
<td>PLR</td>
<td>0.005</td>
<td>0.003</td>
<td>1.005 (0.999-1.011)</td>
<td>0.086</td>
</tr>
<tr>
<td>RPR</td>
<td>2.620</td>
<td>12.244</td>
<td>13.738 (0.00-363400)</td>
<td>0.831</td>
</tr>
<tr>
<td>MLR</td>
<td>0.340</td>
<td>0.967</td>
<td>1.405 (0.211-9339)</td>
<td>0.725</td>
</tr>
<tr>
<td>MPV to PLT</td>
<td>4.471</td>
<td>12.143</td>
<td>87.422 (0.00-189700)</td>
<td>0.713</td>
</tr>
</tbody>
</table>

OR= odds ratio; CI= confidence interval; NLR= neutrophil to lymphocyte ratio; LYM= lymphocyte; PLT= platelet; MPV= mean platelet volume; PLR= platelet to lymphocyte ratio; RPR= red cell distribution width to platelet; MLR= monocyte to lymphocyte ratio; MPV to PLT= mean platelet volume to platelet count ratio.